

# Call Variants with SAMtools Element

Calls SNPs and INDELS with SAMtools mpileup and bcftools.

## Parameters in GUI

Parameter	Description	Default value
<b>Illumina-1.3 + encoding</b>	Assume the quality is in the Illumina 1.3+ encoding (mpileup)(-6).	False
<b>Count anomalous read pairs</b>	Do not skip anomalous read pairs in variant calling (mpileup)(-A).	False
<b>Disable BAQ computation</b>	Disable probabilistic realignment for the computation of base alignment quality (BAQ). BAQ is the Phred-scaled probability of a read base being misaligned. Applying this option greatly helps to reduce false SNPs caused by misalignments (mpileup)(-B).	False
<b>Mapping quality downgrading coefficient</b>	Coefficient for downgrading mapping quality for reads containing excessive mismatches. Given a read with a phred-scaled probability q of being generated from the mapped position, the new mapping quality is about $\sqrt{(\text{INT}-q)/\text{INT}} \times \text{INT}$ . A zero value disables this functionality; if enabled, the recommended value for BWA is 50 (mpileup)(-C).	0
<b>Max number of reads per input BAM</b>	At a position, read maximally the number of reads per input BAM (mpileup)(-d).	250
<b>Extended BAQ computation</b>	Extended BAQ computation. This option helps sensitivity especially for MNPs, but may hurt specificity a little bit (mpileup)(-E).	False
<b>BED or position list file</b>	BED or position list file containing a list of regions or sites where pileup or BCF should be generated. (mpileup)(-I).	
<b>Pileup region</b>	Only generate pileup in region STR (mpileup)(-r).	
<b>Minimum mapping quality</b>	Minimum mapping quality for an alignment to be used (mpileup)(-q).	0
<b>Minimum base quality</b>	Minimum base quality for a base to be considered (mpileup)(-Q).	13
<b>Gap extension error</b>	Phred-scaled gap extension sequencing error probability. Reducing INT leads to longer indels (mpileup)(-e).	20
<b>Homopolymer errors coefficient</b>	Coefficient for modeling homopolymer errors. Given an l-long homopolymer run, the sequencing error of an indel of size s is modeled as $\text{INT} \times s/l$ . (mpileup)(-h).	100
<b>No INDELS</b>	Do not perform INDEL calling (mpileup)(-I).	False
<b>Max INDEL depth</b>	Skip INDEL calling if the average per-sample depth is above INT (mpileup)(-L).	250
<b>Gap open error</b>	Phred-scaled gap open sequencing error probability. Reducing INT leads to more indel calls (mpileup)(-o).	40
<b>List of platforms for indels</b>	Comma delimited list of platforms (determined by @RG-PL) from which indel candidates are obtained. It is recommended to collect indel candidates from sequencing technologies that have low indel error rate such as ILLUMINA. (mpileup)(-P).	
<b>Retain all possible alternate</b>	Retain all possible alternate alleles at variant sites. By default, the view command discards unlikely alleles. (bcf view)(-A).	False
<b>Indicate PL</b>	Indicate PL is generated by r921 or before (ordering is different) (bcf view)(-F).	False
<b>No genotype information</b>	Suppress all individual genotype information (bcf view)(-G).	False
<b>A/C/G/T only</b>	Skip sites where the REF field is not A/C/G/T (bcf view)(-N).	False

<b>List of sites</b>	List of sites at which information are outputted (bcf view)(-l).	
<b>QCALL likelihood</b>	Output the QCALL likelihood format (bcf view)(-Q).	False
<b>List of samples</b>	List of samples to use. The first column in the input gives the sample names and the second gives the ploidy, which can only be 1 or 2. When the 2nd column is absent, the sample ploidy is assumed to be 2. In the output, the ordering of samples will be identical to the one in FILE (bcf view)(-s).	
<b>Min samples fraction</b>	skip loci where the fraction of samples covered by reads is below FLOAT (bcf view)(-d).	0
<b>Per-sample genotypes</b>	Call per-sample genotypes at variant sites. (bcf view)(-g).	True
<b>INDEL-to-SNP Ratio</b>	Ratio of INDEL-to-SNP mutation rate. (bcf view)(-i).	-1
<b>Max P(ref D)</b>	A site is considered to be a variant if P(ref D)	0.5
<b>Prior allele frequency spectrum</b>	If STR can be full, cond2, flat or the file consisting of error output from a previous variant calling run (bcf view)(-P).	full
<b>Mutation rate</b>	Scaled mutation rate for variant calling (bcf view)(-t).	0.001
<b>Pair/trio calling</b>	Enable pair/trio calling. For trio calling, option -s is usually needed to be applied to configure the trio members and their ordering. In the file supplied to the option -s, the first sample must be the child, the second the father and the third the mother. The valid values of STR are pair, trioauto, trioxd and trioxs, where pair calls differences between two input samples, and trioxd (trioxs) specifies that the input is from the X chromosome non-PAR regions and the child is a female (male) (bcf view)(-T).	
<b>N group-1 samples</b>	Number of group-1 samples. This option is used for dividing the samples into two groups for contrast SNP calling or association test. When this option is in use, the following VCF INFO will be outputted: PC2, PCHI2 and QCHI2 (bcf view)(-1).	0
<b>N permutations</b>	Number of permutations for association test (effective only with -1) (bcf view)(-U).	0
<b>Min P (chi^2)</b>	Only perform permutations for P(chi^2).	0.01
<b>Minimum RMS quality</b>	Minimum RMS mapping quality for SNPs (varFilter) (-Q).	10
<b>Minimum read depth</b>	Minimum read depth (varFilter) (-d).	2
<b>Maximum read depth</b>	Maximum read depth (varFilter) (-D).	10000000
<b>Alternate bases</b>	Minimum number of alternate bases (varFilter) (-a).	2
<b>Gap size</b>	SNP within INT bp around a gap to be filtered (varFilter) (-w).	3
<b>Window size</b>	Window size for filtering adjacent gaps (varFilter) (-W).	10
<b>Strand bias</b>	Minimum P-value for strand bias (given PV4) (varFilter) (-1).	0.0001
<b>BaseQ bias</b>	Minimum P-value for baseQ bias (varFilter) (-2).	1e-100
<b>MapQ bias</b>	Minimum P-value for mapQ bias (varFilter) (-3).	0
<b>End distance bias</b>	Minimum P-value for end distance bias (varFilter) (-4).	0.0001
<b>HWE</b>	Minimum P-value for HWE (plus F).	0.0001
<b>Log filtered</b>	Print filtered variants into the log (varFilter) (-p).	False

## Parameters in Workflow File

**Type:** call\_variants

Parameter	Parameter in the GUI	Type
illumina13-encoding	Illumina-1.3+ encoding	<i>boolean</i>
use_orphan	Count anomalous read pairs	<i>boolean</i>
disable_baq	Disable BAQ computation	<i>boolean</i>
capq_thres	Mapping quality downgrading coefficient	<i>numeric</i>
max_depth	Max number of reads per input BAM	<i>numeric</i>
ext_baq	Extended BAQ computation	<i>boolean</i>
bed	BED or position list file	<i>string</i>
reg	Pileup region	<i>string</i>
min_mq	Minimum mapping quality	<i>numeric</i>
min_baseq	Minimum base quality	<i>numeric</i>
extQ	Gap extension error	<i>numeric</i>
tandemQ	Homopolymer errors coefficient	<i>numeric</i>
no_indel	No INDELs	<i>boolean</i>
max_indel_depth	Max INDEL depth	<i>numeric</i>
openQ	Gap open error	<i>numeric</i>
pl_list	List of platforms for indels	<i>string</i>
keepalt	Retain all possible alternate	<i>boolean</i>
fix_pl	Indicate PL	<i>boolean</i>
no_genotype	No genotype information	<i>boolean</i>
acgt_only	A/C/G/T only	<i>boolean</i>
bcf_bed	List of sites	<i>string</i>
qcall	QCALL likelihood	<i>boolean</i>
samples	List of samples	<i>string</i>
min_smpl_frac	Min samples fraction	<i>numeric</i>
call_gt	Per-sample genotypes	<i>boolean</i>
indel_frac	INDEL-to-SNP Ratio	<i>numeric</i>
pref	Max P(ref D)	<i>numeric</i>
ptype	Prior allele frequency spectrum	<i>string</i>
theta	Mutation rate	<i>numeric</i>
ccall	Pair/trio calling	<i>string</i>
n1	N group-1 samples	<i>numeric</i>
n_perm	N permutations	<i>numeric</i>
min_perm_p	Min P(chi^2)	<i>numeric</i>
min-qual	Minimum RMS quality	<i>numeric</i>
min-dep	Minimum read depth	<i>numeric</i>
max-dep	Maximum read depth	<i>numeric</i>
min-alt-bases	Alternate bases	<i>numeric</i>

gap-size	Gap size	<i>numeric</i>
window"	Window size	<i>numeric</i>
min-strand	Strand bias	<i>numeric</i>
min-baseQ	BaseQ bias	<i>string</i>
min-mapQ	MapQ bias	<i>numeric</i>
min-end-distance	End distance bias	<i>numeric</i>
min-hwe	HWE	<i>numeric</i>
print-filtered	Log filtered	<i>boolean</i>

## Input/Output Ports

The element has 2 *input ports*:

**Name in GUI:** Input assembly

**Name in Workflow File:** in-assembly

**Slots:**

Slot In GUI	Slot in Workflow File	Type
Dataset name	dataset	<i>string</i>
Source url	url	<i>string</i>

**Name in GUI:** Input sequences

**Name in Workflow File:** in-sequence

**Slots:**

Slot In GUI	Slot in Workflow File	Type
Source url	url	<i>string</i>

And 1 *output port*:

**Name in GUI:** Output variations

**Name in Workflow File:** out-variations

**Slots:**

Slot In GUI	Slot in Workflow File	Type
Variation track	variation-track	<i>variation</i>